



CHI (RI)CERCA TROVA

UN CICLO DI **WEBINAR** PER CONOSCERE
I **RISULTATI DEI PROGETTI AGER**

30 Aprile 2021

Qualità e sostenibilità economica del pesce allevato



*Fine Feed For Fish è un progetto sostenuto da **AGER - AGroalimentare E Ricerca**, Grant 2016-0101*

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Ordine dei Medici Veterinari della provincia di Milano



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*Lo studio del microbiota intestinale, per la formulazione dei mangimi:
manipolazione delle comunità microbiche intestinali del pesce in allevamento intensivo*

Simona Rimoldi, Genciana Terova

simona.rimoldi@uninsubria.it

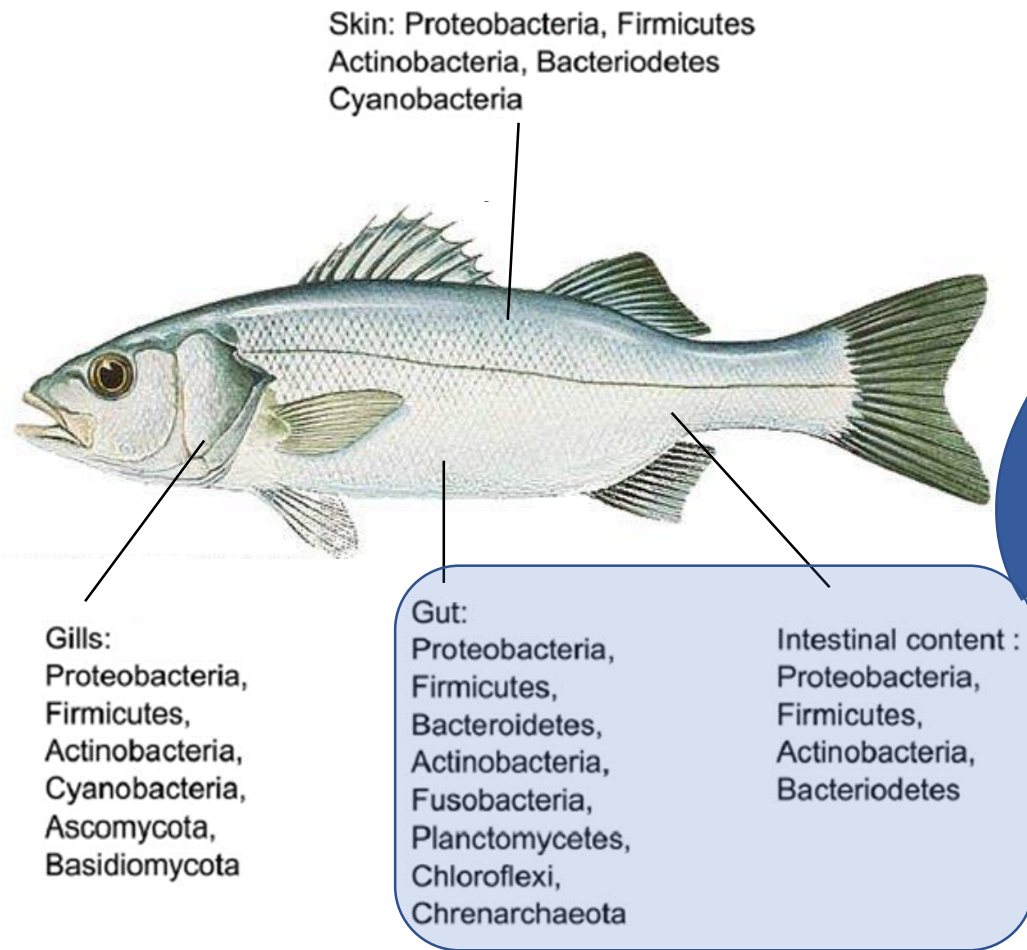
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Fine Feed For Fish è un progetto sostenuto da AGER - AGroalimentare E Ricerca, Grant 2016-0101

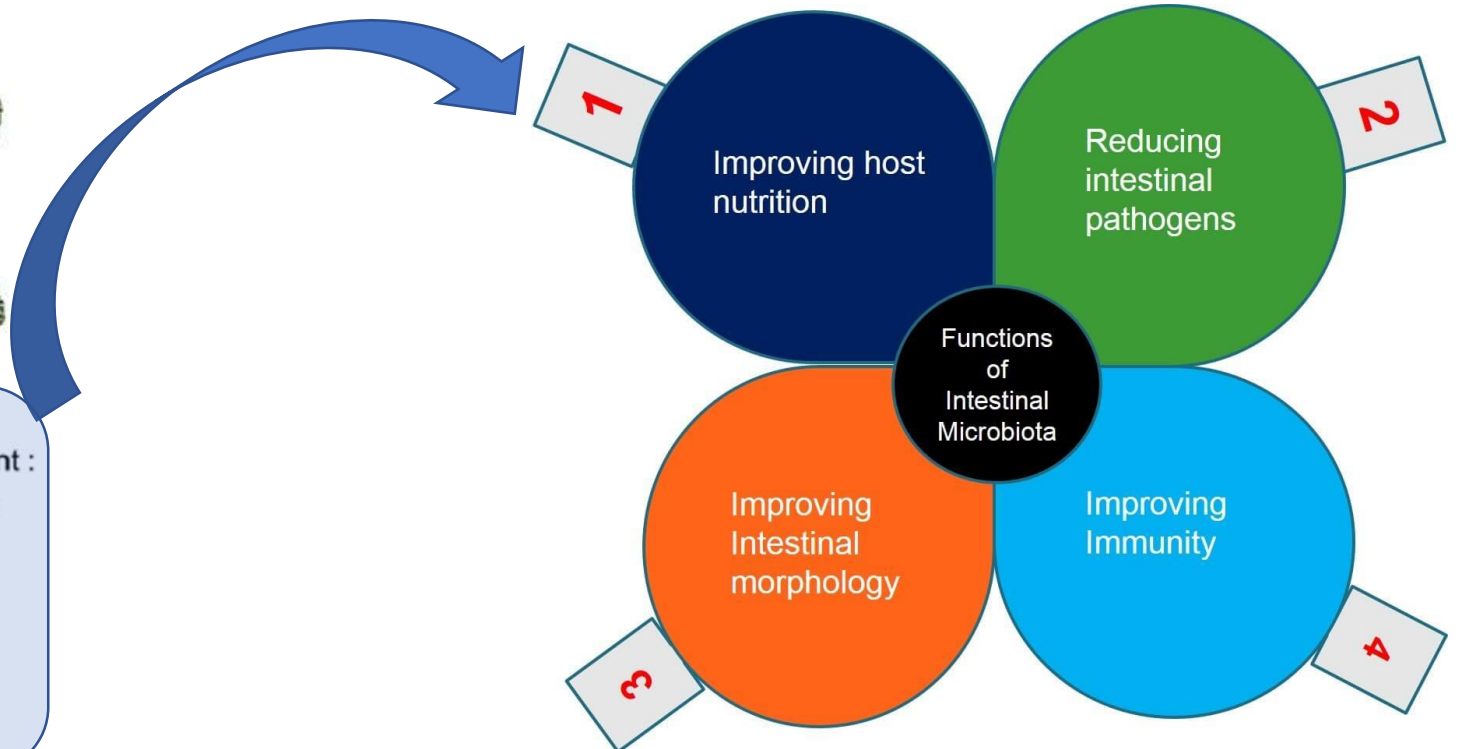


Fish microbiota overview

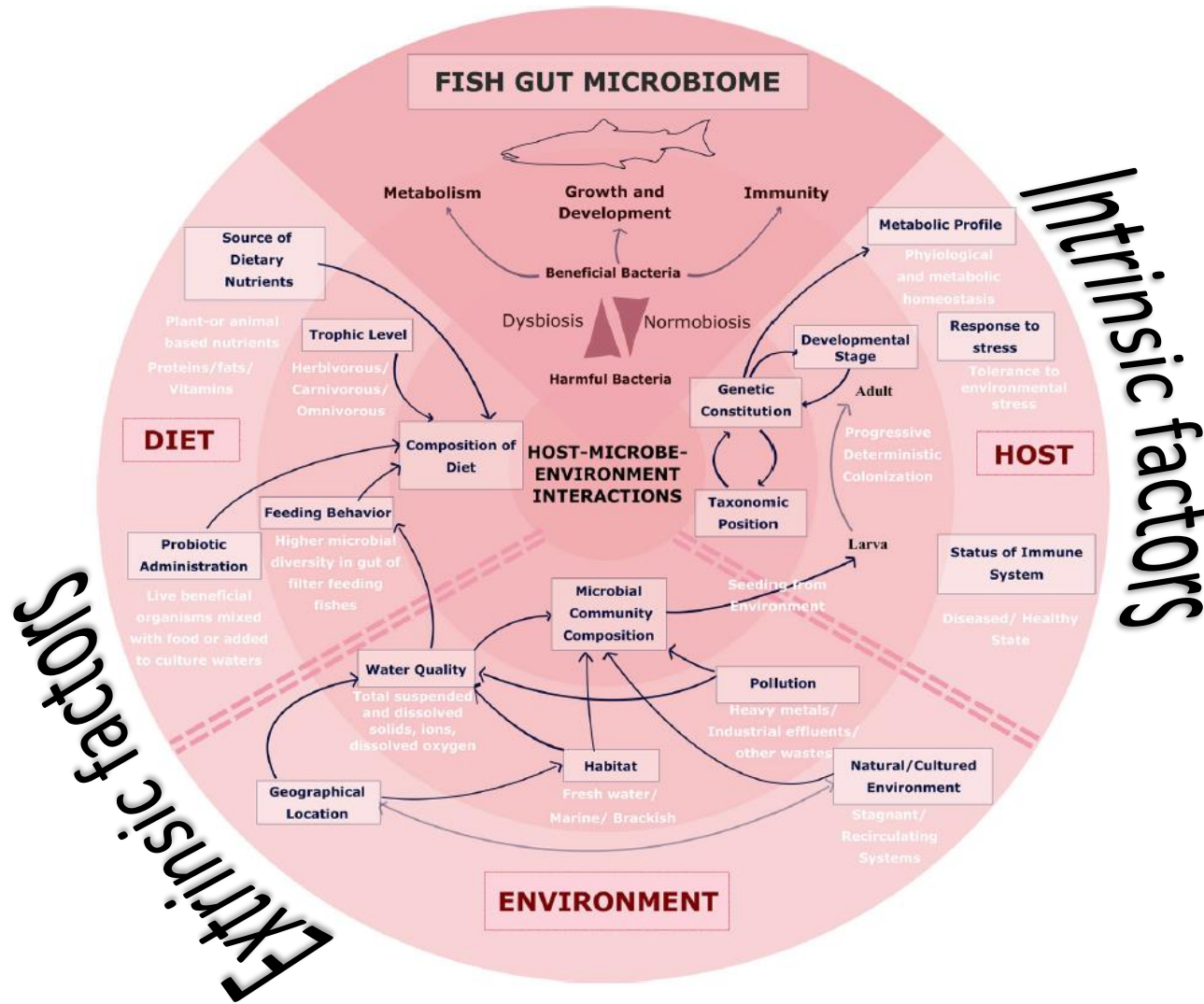
Fish microbiota is a complex community comprising bacteria, fungi, and viruses, inhabits the skin, gills, and gastrointestinal tract.



Gut microbiota balance is essential to maintain overall health in fish
(growth performances, feed efficiency, energy uptake, feed conversion rate and resistance to diseases)



Factors influencing the diversity and function of the gut microbiota of fish.



The imbalance between beneficial and harmful microorganisms (**Dysbiosis**) has deleterious impacts to the gut, including impaired digestion, inflammation in the intestinal wall and lack of local intestinal immunity.

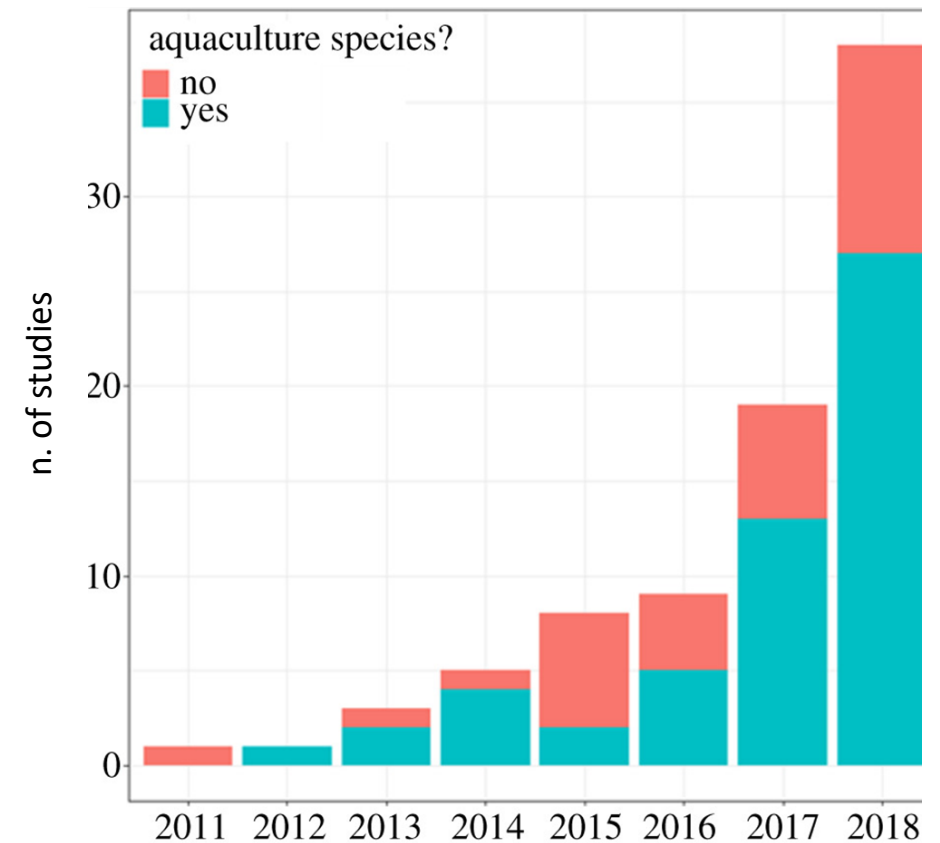


The aquaculture industry is interested in the **manipulation of gut microbiota to improve welfare and nutrition of cultured fish.**

DIET is a major factor driving the composition and metabolism of the gut microbiota.

Different components of the diet can shape the gut bacterial communities

- **PROTEIN AND LIPID SOURCES**
- **PROBIOTICS**
- **PREBIOTICS**
- **BIOACTIVE COMPOUNDS** (organic acids, phytogenics, etc.....)



Perry WB, et al. (2020).Proc. R. Soc. B 287: 20200184



Our research in the project

Evaluate the benefits of novel and more sustainable feed formulations on gut microbiota of cultured fish (European sea bass, gilthead sea bream, and rainbow trout)



Rev Fish Biol Fisheries (2019) 29:465–486
<https://doi.org/10.1007/s11160-019-09558-y>

ORIGINAL RESEARCH

Rainbow trout (*Oncorhynchus mykiss*) gut microbiota is modulated by insect meal from *Hermetia illucens* prepupae in the diet

Genciana Terova¹ · Simona Rimoldi¹ · Chiara Ascione¹ · Elisabetta Gini¹ · Chiara Cecot¹ · Laura Gasco¹

Fish Physiol Biochem
<https://doi.org/10.1007/s10695-020-00918-1>

Intestinal microbial communities of rainbow trout (*Oncorhynchus mykiss*) may be improved by feeding a *Hermetia illucens* meal/low-fishmeal diet

Simona Rimoldi¹ · Mikaela Antonini¹ · Laura Gasco¹ · Federico Moroni¹ · Genciana Terova¹



Article

The Effects of Dietary Insect Meal from *Hermetia illucens* Prepupae on Autochthonous Gut Microbiota of Rainbow Trout (*Oncorhynchus mykiss*)

Simona Rimoldi¹ · Elisabetta Gini¹ · Federica Iannini¹ · Laura Gasco¹ · Genciana Terova^{1,2}

frontiers
in Marine Science

ORIGINAL RESEARCH
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OPEN ACCESS

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Fotini Kokkou,
Wageningen University and Research,

Federico Moroni^{1*}, Fernando Naya-Català^{2*}, M. Carla Piazzon³, Simona Rimoldi¹, Josep Calduch-Giner², Alberto Giardini¹, Inés Martínez², Fabio Brambilla⁴, Jaume Pérez-Sánchez² and Genciana Terova^{1*}



Rimoldi et al. BMC Veterinary Research (2020) 16:118
<https://doi.org/10.1186/s12917-020-02235-1>

BMC Veterinary Research

RESEARCH ARTICLE

Open Access

Effects of hydrolyzed fish protein and autolyzed yeast as substitutes of fishmeal in the gilthead sea bream (*Sparus aurata*) diet, on fish intestinal microbiome

S. Rimoldi¹, E. Gini¹, J. F. A. Koch², F. Iannini¹, F. Brambilla³ and G. Terova^{1*}



PLOS ONE

RESEARCH ARTICLE

Assessment of dietary supplementation with galactomannan oligosaccharides and phytogenics on gut microbiota of European sea bass (*Dicentrarchus Labrax*) fed low fishmeal and fish oil based diet

Simona Rimoldi^{1*}, Silvia Torrealba^{2*}, Daniel Montero², Elisabetta Gini¹, Alex Mako³, Victoria Valdegreño V.⁴, Maitel Iquiedo⁵, Genciana Terova^{1,6*}



Effect of a specific composition of short- and medium-chain fatty acid 1-Monoglycerides on growth performances and gut microbiota of gilthead sea bream (*Sparus aurata*)

Simona Rimoldi¹, Emt Gilozhen¹, Chiara Ascione¹, Elisabetta Gini¹ and Genciana Terova^{1,2}

Terova et al. Journal of Animal Science and Biotechnology
<https://doi.org/10.1186/s13041-021-00101-6>

Journal of Animal Science and
Biotechnology

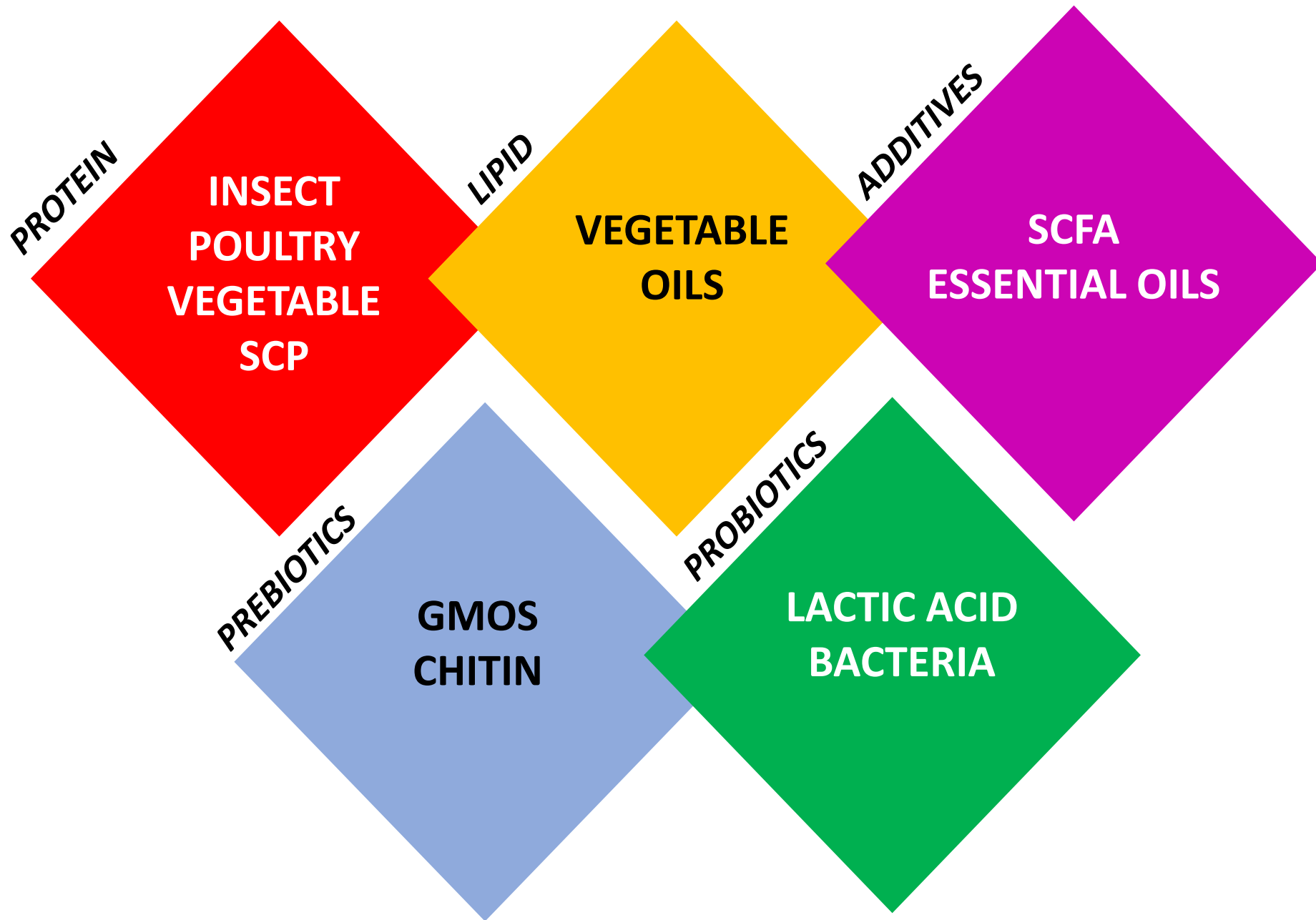
RESEARCH

Open Access

Effects of full replacement of dietary fishmeal with insect meal from *Tenebrio molitor* on rainbow trout gut and skin microbiota

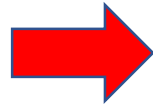
Genciana Terova^{1*}, Elisabetta Gini¹, Laura Gasco¹, Federico Moroni¹, Mikaela Antonini¹ and Simona Rimoldi¹





INSECT MEAL

Black Soldier Fly
(*Hermetia illucens*)

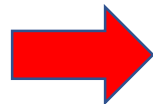


Rainbow trout (*Oncorhynchus mykiss*)

Gilthead sea bream (*Sparus aurata*)



Yellow Mealworm
(*Tenebrio molitor*)



Rainbow trout (*Oncorhynchus mykiss*)

INSECT MEAL

Hermetia illucens larva meal was provided by MUTATEC (Caumont-sur-Durance, France)

<i>H. illucens</i> Meal	DIET			
	Hi 0	Hi 10	Hi 20	Hi 30
Ingredients (% as it)				
Fishmeal ^a	60	54	48	42
<i>Hermetia illucens</i> meal	0	10	20	30
Fish oil	7	7	7	7
Soybean oil	5	4	3	2
Wheat bran	10	7	4	1
Wheat meal	4	4	4	4
Starch gelatinized D500	11	11	11	11
Vitamin premix ^b	1.5	1.5	1.5	1.5
Mineral premix ^c	1.5	1.5	1.5	1.5

Ingredients (g Kg ⁻¹)	Ctrl	Hi15
Fishmeal ^a	200.0	100.0
<i>Hermetia illucens</i> larva meal ^b	0.0	150.0
Wheat gluten	130.0	130.0
Soybean meal	200.0	200.0
Porcine haemoglobin	92.0	82.0
Wheat starch	233.9	193.9
Fish oil	69.8	69.8
Soybean oil	69.8	69.8
Minerals ^c	2.5	2.5
Vitamins ^d	2.0	2.0

FISH

INSECT

DIET

RESULTS



Rainbow trout

Hermetia illucens



0, 10, 20, 30%
Inclusion

15% inclusion
(50% FM replacement)

- SGR and WG were comparable among the treatments. FCR was < 1 in all groups
- Hi meal inclusion ($\geq 15\%$) in the diet positively modulated both allochthonous and autochthonous gut microbiota, increasing species richness and diversity
- Several bacteria taxa were discriminatory between diets

Allochthonous microbiota:

- ❑ the amounts of **Actinobacteria** and **Firmicutes** were positively modulated by Hi. Proteobacteria decreased with Hi dietary inclusion
- ❑ fish fed with Hi meal had **higher percentage of Actinomyces, LAB** (*Facklamia*, *Enterococcus*, *Lactobacillus*, and *Pediococcus*), **and Bacillus**. **Many beneficial bacterial species were correlated to Hi, some of them were butyrate producers** (*Corynebacterium variabile*, *Lactobacillus paraplantarum*, *Lactobacillus zae*, *Weissella cibaria*, *Clostridium butyricum*, and *Clostridium fimentarium*)

Autochthonous microbiota:

- ❑ **Tenericutes** and **Proteobacteria** were significantly influenced by insect meal inclusion (20% and 30%) in the diet
- ❑ trout fed Hi 20 and Hi 30 diets were enriched in *Mycoplasma* genus
- ❑ Hi meal caused a significant reduction of **Proteobacteria** both in the gut digesta and mucosa. **All genera adversely affected by insect-meal diets (*Shewanella*, *Aeromonas*, *Citrobacter*) were Gram-negative bacteria that include potential pathogen species.**

- Dietary Hi meal upregulated those pathways involved in starch and sugar metabolism

Terova et al. (2019). *Rev Fish Biol Fisheries* 29:465–486

Rimoldi et al. (2019). *Animals* 9, 143

Rimoldi et al. (2021) *Fish Physiol Biochem* 47(2):365-380



INSECT MEAL



	DIET	
	Hi 0	Hi 10
Fishmeal	15.0	5.0
Processed animal protein from poultry	10.7	10.7
SCP	3.9	3.9
<i>Hermetia illucens</i> meal*	0	10.0
Vegetable meals	34	34
Maize gluten meal,	18.9	18.9
Fish oil	3.9	5.0
Rapeseed oil	9.1	7.8
Vitamin & mineral premix	1.15	1.15

*Insect meal was provided by HIPROMINE (Poland)



Item	TM 0	TM 100
Ingredients, %		
Fishmeal 65 (Peruvian)	20	-
<i>Tenebrio molitor</i> larvae meal	-	20
Soy protein concentrate	18	18
Wheat gluten	7.75	7.06
Corn gluten	8	8
Soybean meal (48%)	7	7
Wheat meal	14.23	13.8
Sardine oil	4.3	4.1
Soybean oil	8.6	8.2
Rapeseed oil	8.6	8.2
Soy lecithin	0.5	0.5
Vitamin ^a and mineral premix ^b	1	1

SPAROS LDA
Ÿnsect (Evry, France)

FISH	INSECT	DIET	RESULTS
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Gilthead sea bream

Hermetia illucens



0, 10%
inclusion

- Hi inclusion improved the growth performances and FCR
- No differences in alpha- and beta- diversity
- Gut autochthonous microbial communities were not affected by 10% inclusion of Hi meal


Rainbow trout



Tenebrio molitor

0, 20%
Inclusion
(FM substitution of 0,
100%)

- The growth parameters were comparable between two feeding groups.
- Skin microbiota had higher species richness and biodiversity than intestinal ones.
- No evident sign of dysbiosis, but only slight microbiota changes:
 - Gut mucosa:**
reduction of Proteobacteria, Neisseriaceae and Ruminococcaceae families and *Citrobacter* genus
 - Skin mucosa:**
reduction of Neisseriaceae (g_ *Deefgea*) and increase of Clostridiaceae

Terova et al. Journal of Animal Science and Biotechnology (2021) 12:30



POULTRY MEAL & VEGETABLE MEAL

- ❖ Since June 2013, the European Union has re-authorized the use of non-ruminant animal by-product meal in aquafeeds.

FISH	DIET	RESULTS
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Rainbow trout

Seven diets: 25-69%
Inclusion of alternative animal proteins
CTRL: diet E

Table 2. Formul

	A	B	C	D	E	F	G
Fish meal							
Dried swine hem							
Dried swine plas							
Poultry by-prod							
Fish oil							
Rapeseed meal							
Soybean meal							
Guar germ meal							
Wheat flour							
Corn gluten							
Vital wheat glute							
Peas							
Soy protein conc							
Soybean oil							
DL- methionine							
Lisin							
Taurin							
Antioxidants pre							
Vitamin and mir							
Stay C 35%							
FP/TP	20.6	20.9	10.6	11.2	37.3	20.0	11.0
TAP/TP	68.0	68.0	64.0	56.0	62.0	75.0	80.0
AP (TAP-FP)	47.4	47.4	53.4	44.8	24.7	55.0	69.0
VP/TP	32.0	32.0	36.0	44.0	38.0	25.0	20.0

- Diet D gave the best SGR and FCR (0.96), while A and B were the worst.
- No overall effect on bacterial richness and diversity in response to FM substitution.
- Beta diversity analysis revealed a relationship between diet type and gut microbiota.
- The inclusion of at least 25% of plant proteins in the diet favored the presence Firmicutes
- Highest percentage of PBM of commercial F and G diets had an adverse effect on the gut microbial community by reducing LAB.
- *Corynebacterium* and *Fusobacterium* genera were enriched in the intestine of D group with the best performances

➤ ***bacteria belonging to Fusobacteria phylum can produce butyrate and synthesize vitamins***

Rimoldi S, (2018). PLoS ONE 13(3): e0193652.



VEGETABLE MEAL

FISH



Rainbow trout

ITA: non-selected

USA: selected for growth on a all plant-based diet (University of Idaho)

DIET

- **A:** COMPLETE FEED FOR TROUT (commercial diet, Naturalleva) containing ANIMAL PROTEINS (poultry, fish, and shrimp meal)
- **B:** ALL PLANT PROTEIN DIET

RESULTS

- USA strain fed with diet A showed higher final weight and better FCR than ITA strain
- Neither diet nor genotype affected gut bacterial α -diversity
- Gut microbial communities were mainly affected by genotype in term of absence/presence of specific genera:



***Cetobacterium* 10 times more abundant in USA strain**

This genus is capable of producing vitamin B-12 and antimicrobial metabolites



POULTRY, VEGETABLE & INSECT MEAL

Large scale trial

FISH	DIET	RESULTS
 Rainbow trout Fattoria del Pesce (Cerano, NO)	CTRL: 27% FM PM: 12% FM, 15% PM Hi: 12% FM, 10% Hi <i>Hermetia illucens</i> meal was provided by MUTATEC (Caumont-sur-Durance, France)	<ul style="list-style-type: none">• Trout fed insect diet showed higher final weight and SGR than other two groups.• No changes in alpha- and beta-diversity• Diet did not affect gut microbial composition
 Gilthead sea bream Croatia	A: 20% PM + 10% FM B: 20% PM + 10% VM C: commercial diet	<ul style="list-style-type: none">• Commercial diet gave the best SGR, while economic FCR was improved by diet A• No changes in alpha- and beta-diversity.• Diet did not affect gut microbial composition.

SCP – yeast protein

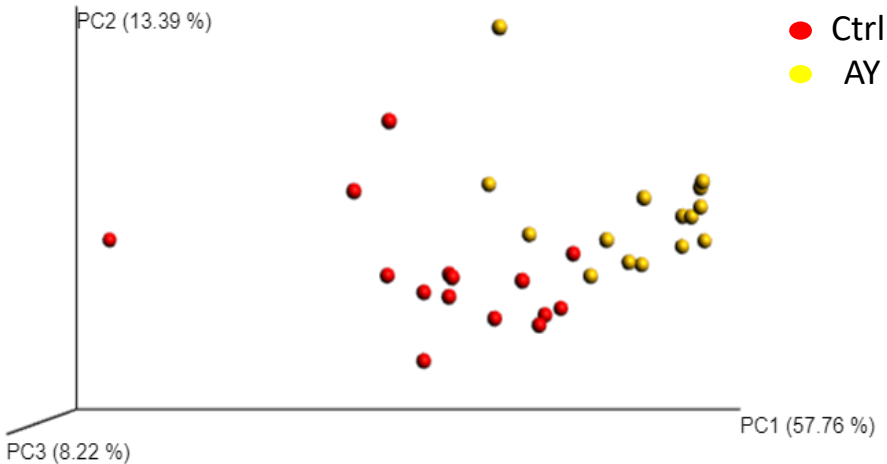
FISH	DIET	RESULTS
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Gilthead sea bream

- AY: 17.8% FM and 4.6% autolysed yeast
- Ctrl: 22% FM

- Dietary inclusion of 5% AY did not affect intestinal bacterial species richness
- Changes in beta-diversity, were found both in type and abundance of specific taxa.
- Indigestible carbohydrate degrading and SCFA producing bacteria, were positively affected by AY
 - ❖ only sea bream fed with AY harbored members of genera *Prevotella* and *Megasphera*
- AY dietary inclusion promoted the growth of *Bacillus* genus.



	DIETS	
	Ctrl	AY
Ingredients (% as it is):		
Fishmeal	22.25	17.80
Corn gluten meal	17.80	17.71
Guar germ meal	15.13	17.68
Soybean meal	10.70	10.68
Soy Protein Concentrate	9.38	10.04
Wheat middling	7.45	7.12
Fish oil 92	6.93	6.93
Fish protein hydrolysed		
HiCell® – autolysed yeast		4.60
Pea meal	4.45	1.53
Cameline oil	2.42	2.42
Mineral/Vitamin supplement	2.00	2.00
Rapeseed oil	1.49	1.49

***HiCell** (Biorigin) is an autolysed dry yeast obtained from the fermentation of a strain of *Saccharomyces cerevisiae* (OGM free)

Rimoldi et al. BMC Veterinary Research (2020) 16:118



CAMELINA OIL



naturAlleva

Feed ingredients (%)	Diets			
	1 (ctrl)	2	3	4
Fish meal	19.4	19.4	19.4	19.4
Soybean meal	10.1	10.1	10.1	10.1
Guar germ meal	12.7	12.7	12.7	12.7
Wheat	14.5	14.5	14.5	14.5
Corn gluten	18.6	18.6	18.6	18.6
Soy protein concentrate	3.8	3.8	3.8	3.8
Fish oil	15.3	12.4	9.3	6.0
Camelina oil	0.0	2.9	6.0	9.3
Vitamins and minerals ^{a,b}	5.6	5.6	5.6	5.6

FISH	DIET	RESULTS
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Gilthead sea bream

- Diet 1, ctrl diet (100% FO)
- Diet 2 (20% CO, 80% FO)
- Diet 3 (40% CO, 60% FO)
- Diet 4 (60% CO, 40% FO)

Parque Científico-Tecnológico Marino (PCTM) of the University of Las Palmas de Gran Canaria

- FCR and SGR were not affected by diet, final weight was significantly lower for fish fed the 60% CO diet.
- α and β -diversity of gut bacteria in both feces and mucosa were not affected by diet
- In gut mucosa *Corynebacterium* and Rhodospirillales (*Acetobacter*, *Enhydrobacter*), were associated with the 60% CO diet

higher abundance of Corynebacterium in the mucosa of fish fed 60% CO diet increased the KEGG pathway of fatty acid synthesis

- In feces, genera of *Peptostreptococcus*, *Vagococcus* and *Massilia* increased in fish fed diets with 60% replacement with CO
- Lowest abundance of LAB, specifically *Lactobacillus*, in the feces of fish fed the 60% CO
- **This study demonstrated that up to 40% of FO can be replaced with CO without negative effects on growth performance, and gut microbiota of gilthead sea bream**

Huyben et al., (2020). PeerJ 8:e10430



PREBIOTICS - CHITIN

FISH



Rainbow trout

RESULTS

- No differences were found between groups for SGR and FCR (1.09 – 1.13)
- Dietary chitin supplementation (1.6%) increased intestinal bacterial species richness and diversity
- Gut microbial community profiles qualitatively differed:
 - ✓ Firmicutes and Actinobacteria were enriched by chitin in the diet
 - ✓ At genus level relative amount of *Corynebacterium* was higher in chitin feeding group – inflammation-inhibiting bacteria genus involved in fatty-acid synthesis
- Specific bacteria genera were found only in trout receiving CHITIN diet:
 - f__ Brevibacteriaceae; g__ Brevibacterium
 - f__ Staphylococcaceae; g__ Jeotgalicoccus
 - f__ Bacillaceae; __g_Bacillus
 - f__ Bacillaceae; __g_Oceanobacillus
 - f__ Actinomycetaceae; g__ Actinomyces
 - f__ Aerococcaceae; g__ Atopostipes
- Dietary CHITIN enhanced SCFA production

Fecal content of SCFA (mM)

Fecal pool	Acetic acid	Propionic acid	Butyric acid
Chitin	16.26	1.22	2.06
Ctrl	6.38	Trace	0.51



GMOS & PHYTOGENICS

Ingredients	Diets (%)			
	CONTROL	GMOS	PHYTO	GMOSPHYTO
Fish meal ¹	10	10	10	10
Soya protein concentrate	18.9	18.9	18.9	18.9
Soya meal	12.0	12.0	12.0	12.0
Corn gluten meal	25.0	25.0	25.0	25.0
Wheat	8.7	8.2	8.7	8.2
Wheat gluten	2.0	2.0	2.0	2.0
Guar meal	8.0	8.0	8.0	8.0
Rapeseed extracted	3.0	3.0	3.0	3.0
Fish oil ²	6.7	6.7	6.7	6.7
Rapeseed oil ³	5.4	5.4	5.4	5.4
Vitamin and mineral premix ⁴	3.7	3.7	3.7	3.7
Antioxidant ⁵	0.06	0.06	0.06	0.06
Galactomannan oligosaccharides ⁶	0	0.5	0	0.5
Phytogenic ⁷	0	0	0.02	0.02

FISH	DIET	RESULTS
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European sea bass

CONTROL: plant-based formulation with 10% FM and 6% FO

GMOS: 0.5%

PHYTO: 0.02% of a mixture of garlic and labiatae-plants oils

GMOSPHYTO: a combination of two 0.52%

- The experimental diets did not induce differences in fish growth or diet utilization.
- The dietary inclusion of GMOS (0.5%) and PHYTO (0.02%) induced changes in bacterial community composition of European sea bass.
- Allochthonous intestinal microbiota of sea bass fed GMOS, but not GMOSPHYTO diet, showed reduced number of Vibrionales.
- the Clostridiales order was more abundant in intestinal content of fish fed PHYTO diet, as well as, in mucosa-associated microbiota of fish receiving GMOSPHYTO diet.

the Clostridiales order includes several butyrate producers. Ruminococcus genus, found only in the intestinal lumen of fish fed PHYTO diet, plays an important role in the degradation of indigestible carbohydrate, thus contributing to the more efficient energy utilization of feed.

- *Escherichia* genus was practically undetectable in allochthonous microbiota of fish fed PHYTO and GMOSPHYTO diets.
- Only GMOSPHYTO diet increased the abundance of Bacteroidales, Lactobacillales, and Clostridiales resident bacterial orders



FISH	DIET	RESULTS
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Gilthead sea bream

CTRL: commercial plant-based diet
Sh108: CTRL + supplemented with 0.5% of SILOhealth 108Z* (SILO SpA)

Rimoldi et al. (2018). PeerJ 6:e5355

- SGR did not reveal any significant differences between control and SILOhealth 108Z-supplemented dietary group. Economic FCR was improved by diet Sh108.
- The relative abundance of Firmicutes was significantly higher in fish fed with diet Sh108 than in controls
- the number of Proteobacteria, in particular Gammaproteobacteria, was negatively affected by SILOhealth 108Z
- The number of bacteria assigned to the *Lactobacillus* genus was significantly higher in Sh108 samples

L. agilis, was only found in fish receiving Sh108 diet

SILOhealth 108Z positively modulated the fish intestinal microbiota by increasing the relative abundance of beneficial lactic acid bacteria



Rainbow trout

CTRL+: fish-based diet
CTRL-: plant-based diet
 CTRL- supplemented with:
0.5%, 1%, or 3% of SILOhealth 108Z* (SILO SpA)

- Alpha and beta diversity analysis did not show any difference
- Fish fed diets supplemented with monobutylin were enriched in Enterobacteriales respect to controls, and showed a decrease in bacteria assigned to *Vibrio* genus
- LAB abundance was not improved by monobutylin inclusion



PROBIOTICS – *Lactococcus lactis* SL242



Gilthead sea bream

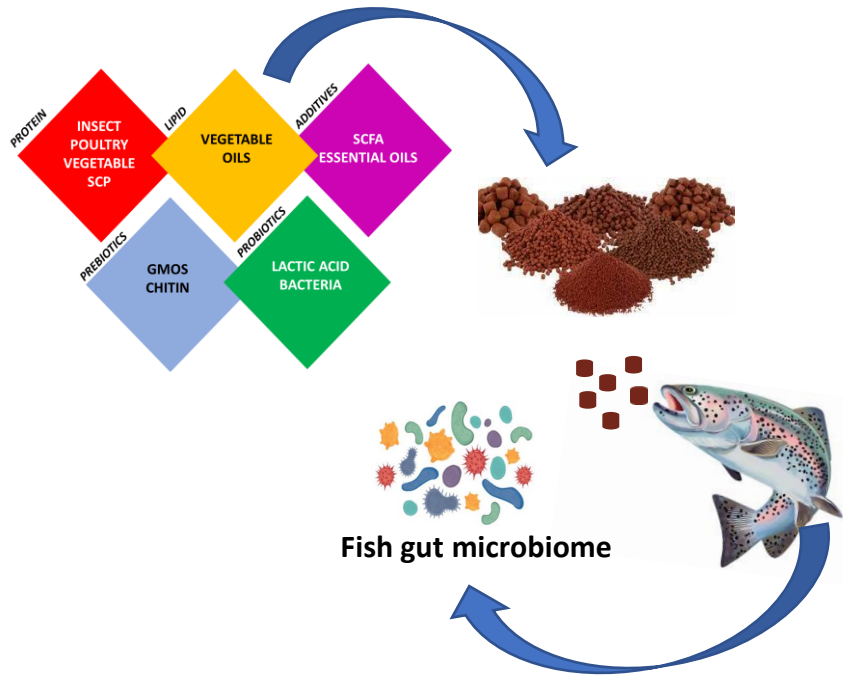
Experimental facilities of IATS-CSIC (Castellón, Spain)

Moroni et al. (2021). *Front. Mar. Sci.* 8:659519

FISH	DIET	RESULTS
 Gilthead sea bream	Diet A: Ctrl	<ul style="list-style-type: none"> There were no significant differences between groups for the FCR and SGR, but final weight of fish fed diet C was higher than control Lack of colonization of the probiotic in the host's intestinal mucosa. Probiotic did modulate the fish gut microbiota: <ul style="list-style-type: none"> bacterial species richness and diversity of group C was lower than control fish fed diet B and C were enriched with Actinomycetales as compared to control. fish fed diets B and C showed a higher Firmicutes/Bacteroidetes ratio than control fish and this could be correlated to better growth performances. group C was characterized by a Proteobacteria/Firmicutes ratio five times higher than in the other groups and higher amount of Spirochetes phylum <ul style="list-style-type: none"> ❖ <i>changes in Proteobacteria/Firmicutes ratio were due to antibiotic nisin A produced by L. lactis SL242</i> pathways related to protein digestion and absorption were over-represented in fish fed with probiotic diets
	Diet B: supplemented with 2.5g / 100kg of <i>L. lactis subsp. lactis</i> (2×10^9 CFU/kg, low dose)	
	Diet C: supplemented with 6.2g / 100Kg of <i>L. lactis subsp. lactis</i> (5×10^9 CFU/Kg, high dose)	

Ingredients	Diet A
Fishmeal	10.1
Corn gluten	24.3
Guar germ meal	10.0
Soybean meal	13.1
Soya protein concentrate	13.6
Wheat	10.8
Fish oil	7.5
Rapeseed oil	3.5
Camelina oil	3.5
Lactic bacteria	0.0
Lysine	0.9
DL-methionine	0.4
Monoammonium phosphate	1.2
Taurine	0.4
Vitamins ^a and Minerals ^b	0.7

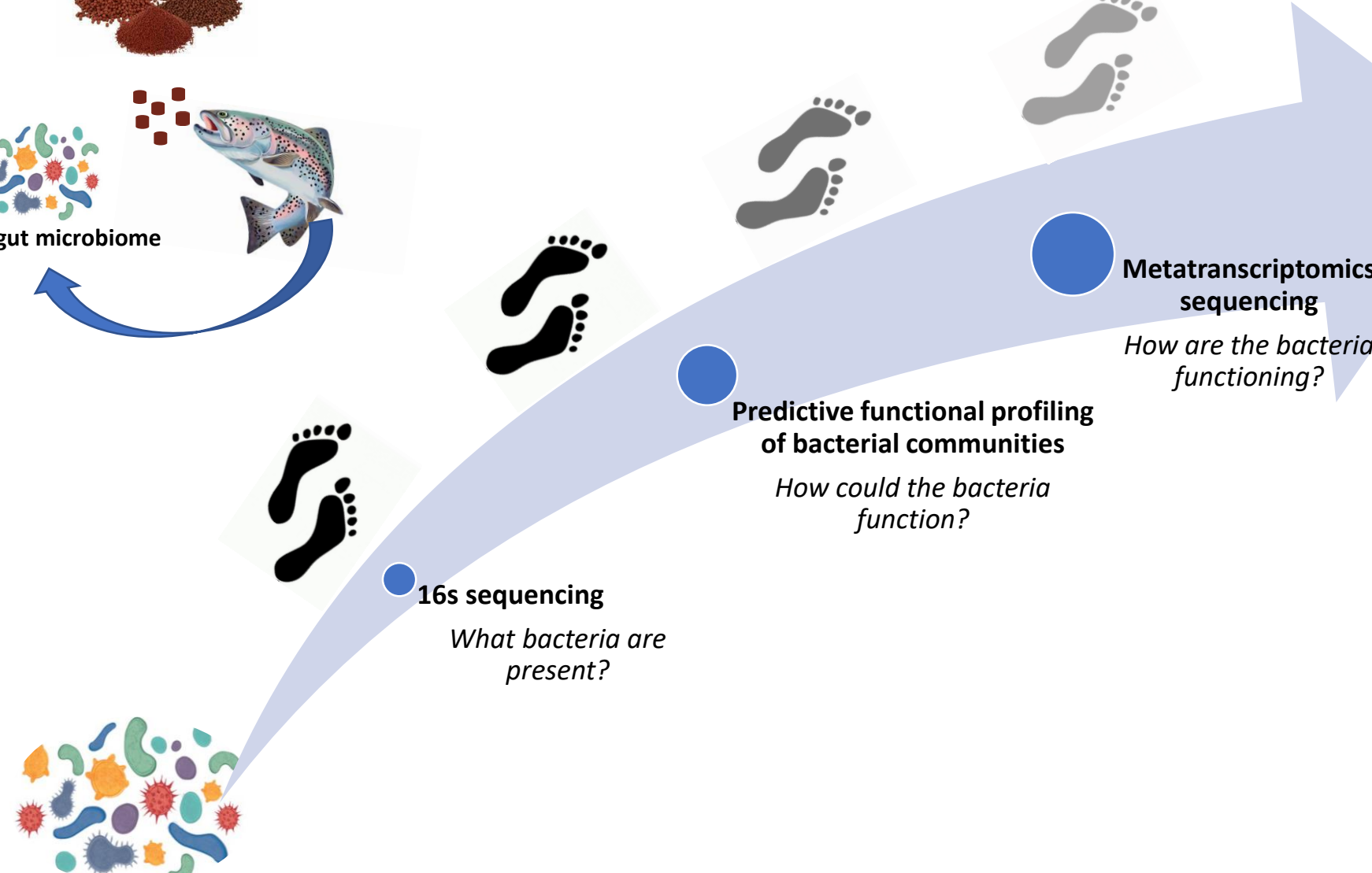




FUTURE STEPS



Selective manipulation of gut microbiota



16s sequencing
What bacteria are present?

Predictive functional profiling of bacterial communities
How could the bacteria function?

Metatranscriptomics sequencing
How are the bacteria functioning?





Prof. Genciana Terova
Prof. Marco Saroglia

Dr. Chiara Ceccotti
Dr. Federico Moroni
Dr. Federica Iannini
Dr. Micaela Antonini
Dr. Simona Rimoldi

thanks